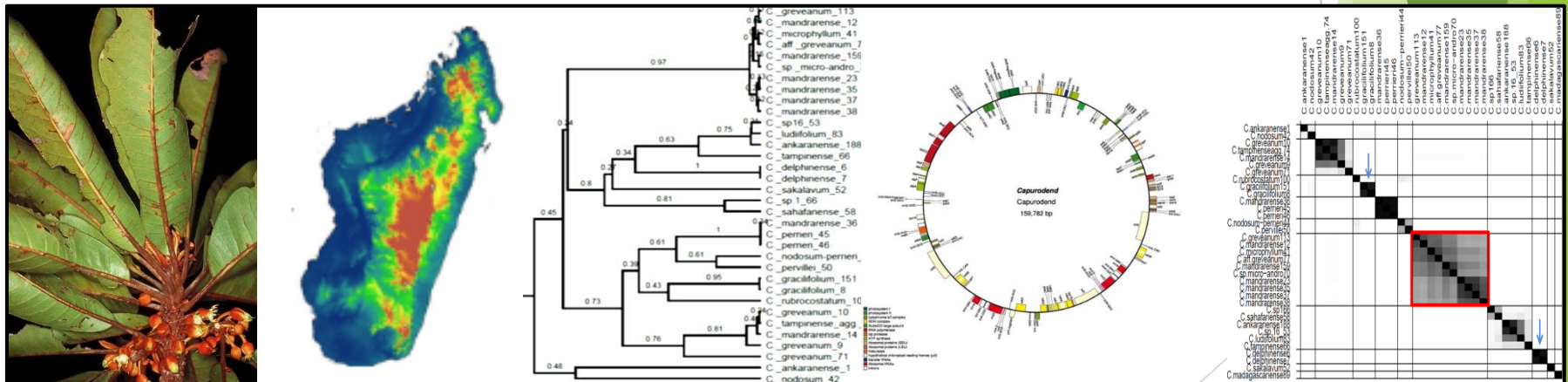


Phylogeny and species delimitation of the Madagascar endemic trees *Capurodendron* using a Gene Capture approach

Carlos G. Boluda

Study supervised by
Yamama Naciri & Laurent Gautier

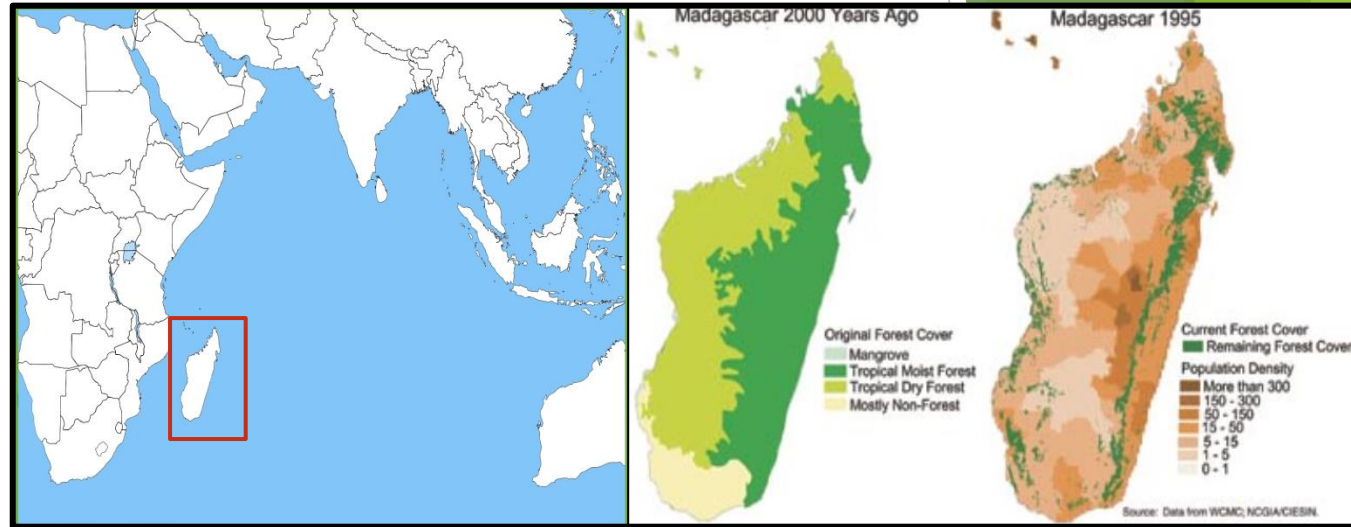


Introduction

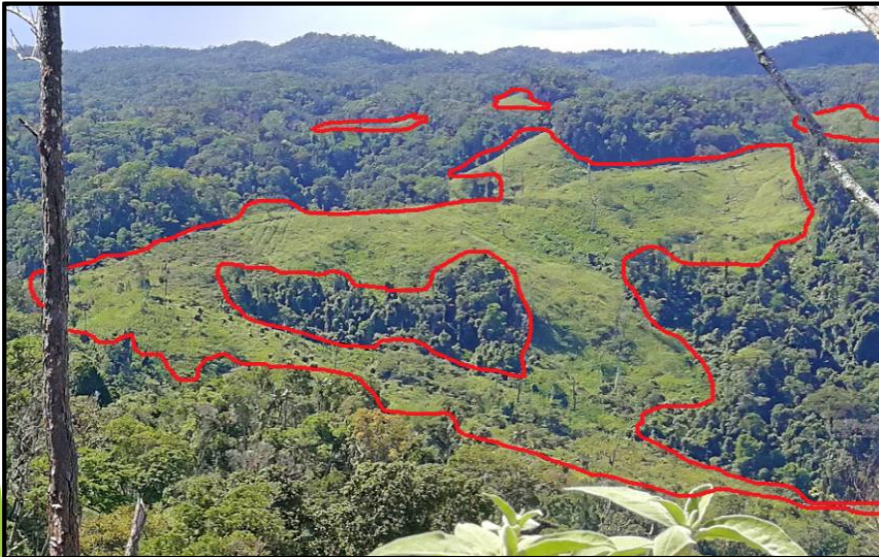
Madagascar:

- Isolated from India ~ 88 mya. (Upper Cretacic).

- 82% of endemic vascular flora.



Deforestation mainly due to:



Slash-and-burn agriculture

Capurodendron also endangered by:

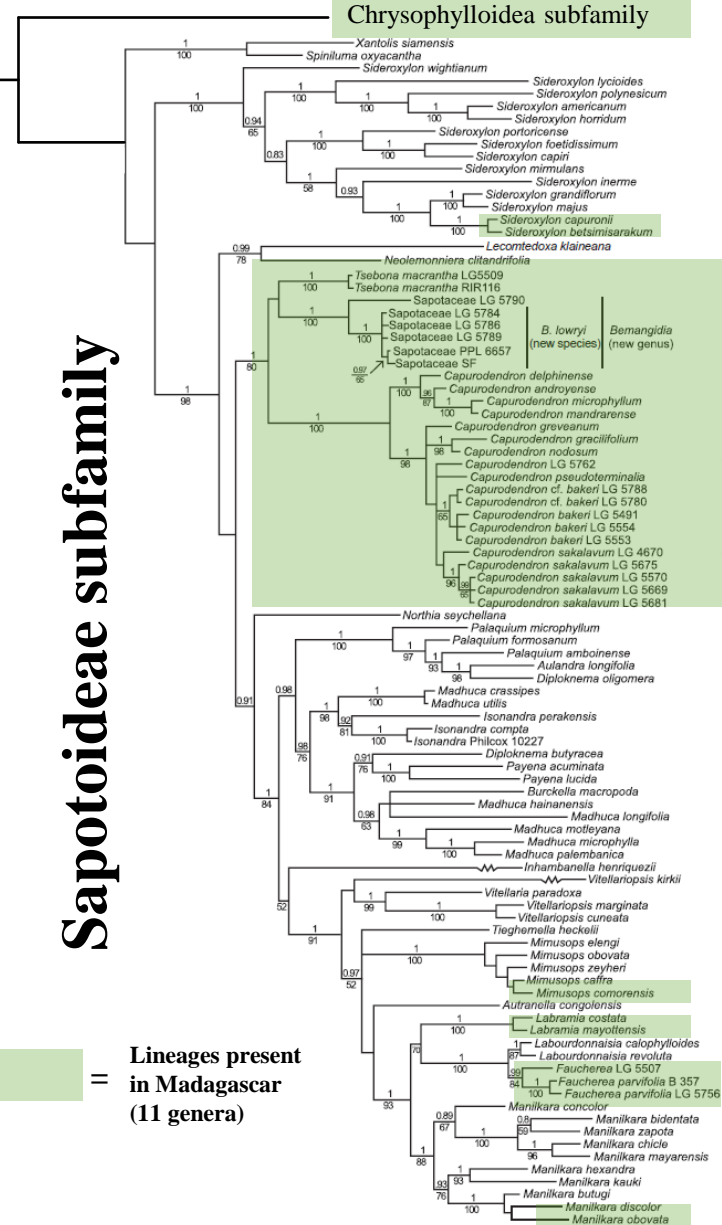


Wood felling

Introduction

Sapotoideae subfamily

Chrysophylloidea subfamily



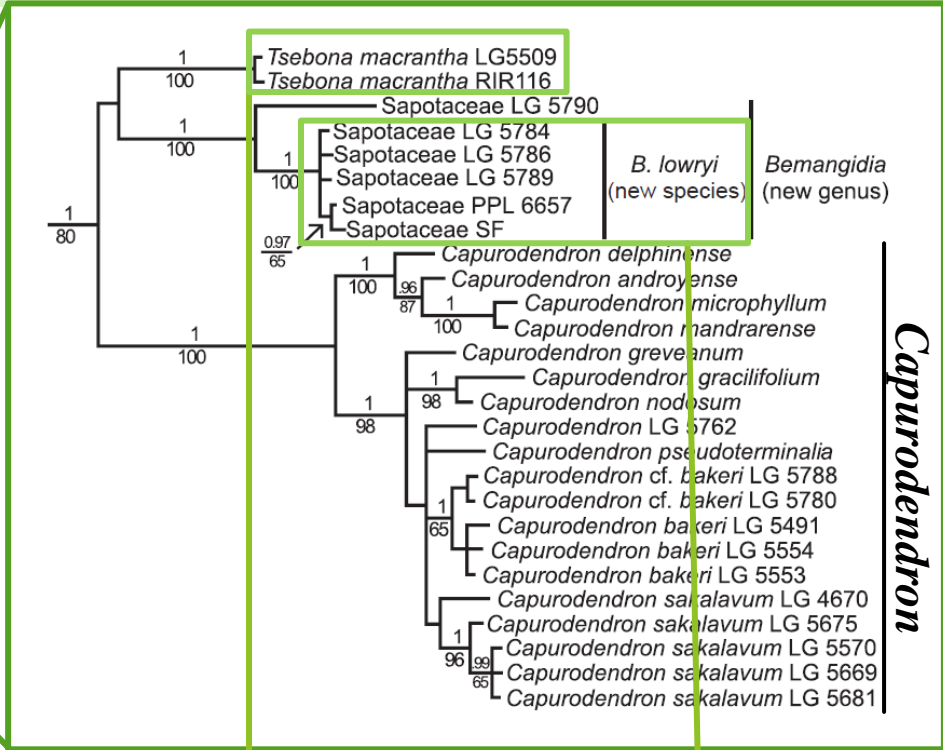
SIDEROXYLEAE

TRIBONEAE (new tribe)

ISONANDREAE

SAPOTEAEE

Tribu Tseboneae



Capurodendron



Tsebona macrantha

Bemangidia lowryi

Lineages present in Madagascar (11 genera)

Introduction

Genus *Capurodendron*:

Undescribed morphologies
Species only known from type specimen
Intermediate morphologies
Morpho/species complexes



Undescribed species
Extinct species
Hybridization
Current speciation



C. delphinense



C. greveanum



C. androyense



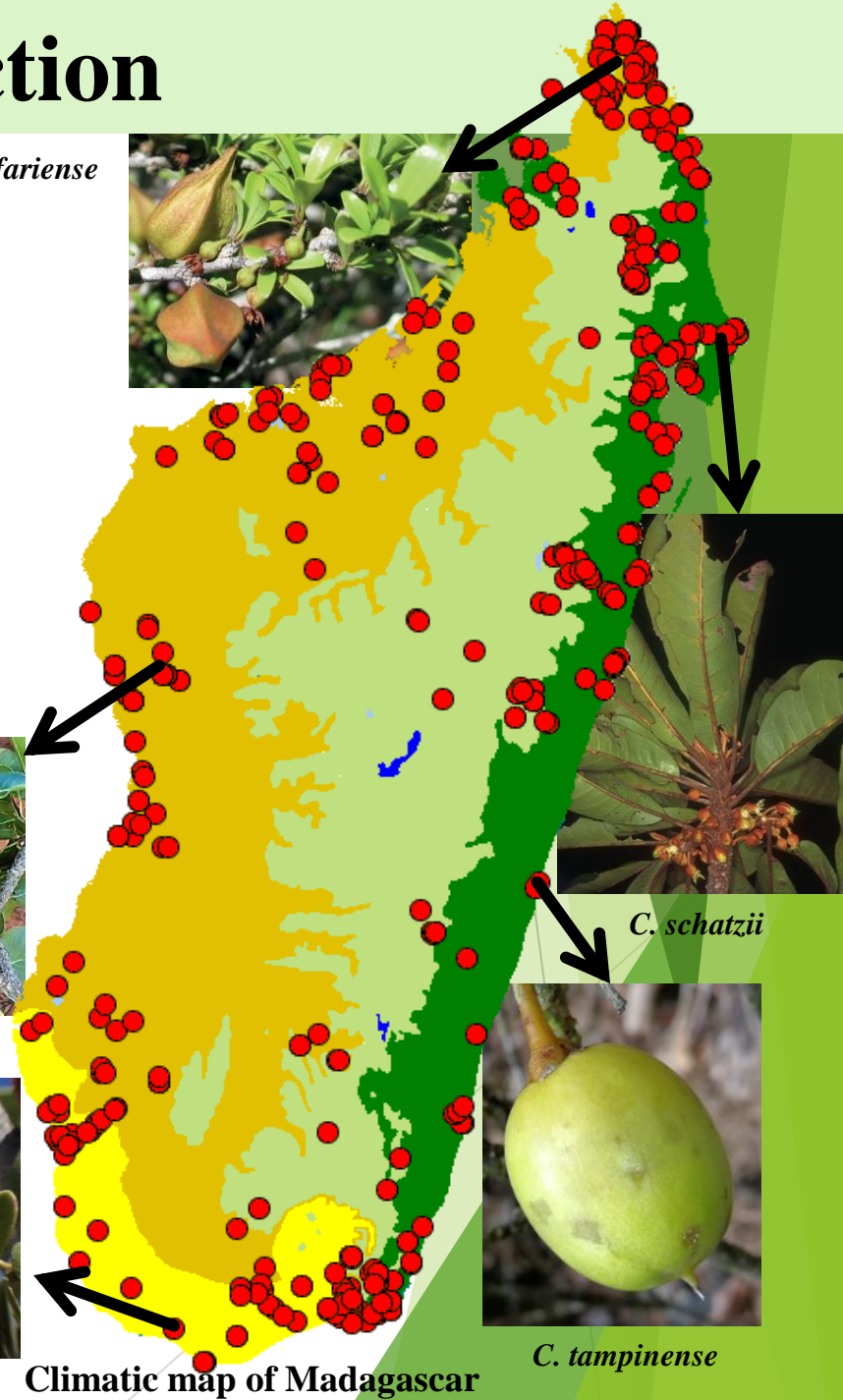
C. sahafariense



C. schatzii



C. tampinense

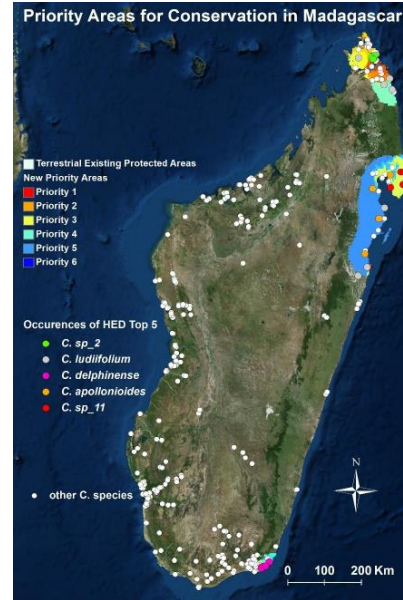


Climatic map of Madagascar

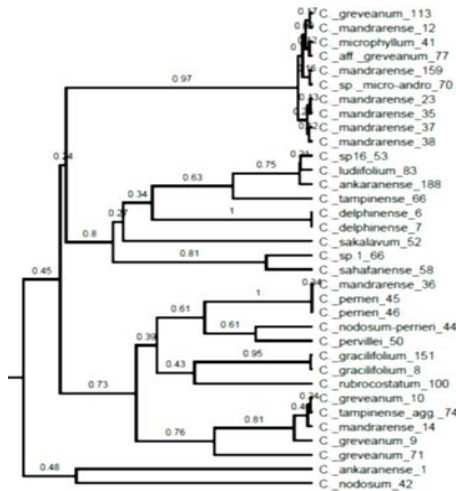
Objectives

Main objectives:

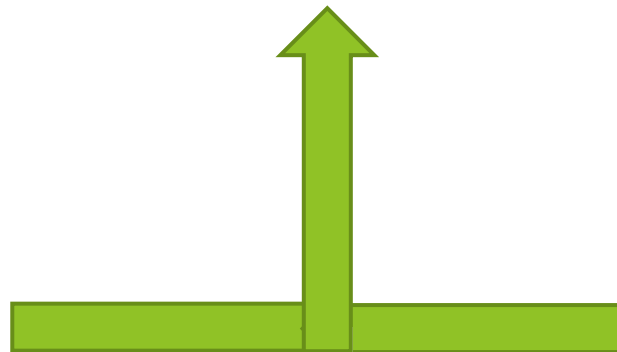
- 1° Delimit the *Capurodendron* species using phylogenomics.
- 2° Estimate the potential distribution of each species and establish the UICN protection categories.



Priority areas for conservation



Species delimitation



Potential distribution, UICN category

Materials and Methods

Sequences obtention:

Hundreds of genes per specimen are wanted

Undescribed species
Extinct species
Hybridization
Current speciation



Silica gel samples



Herbarium samples up to 80 years old



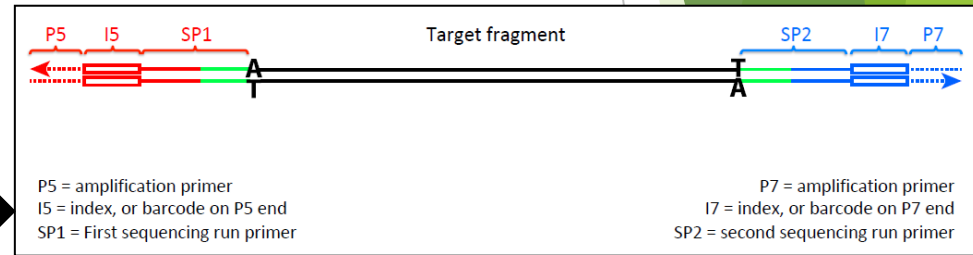
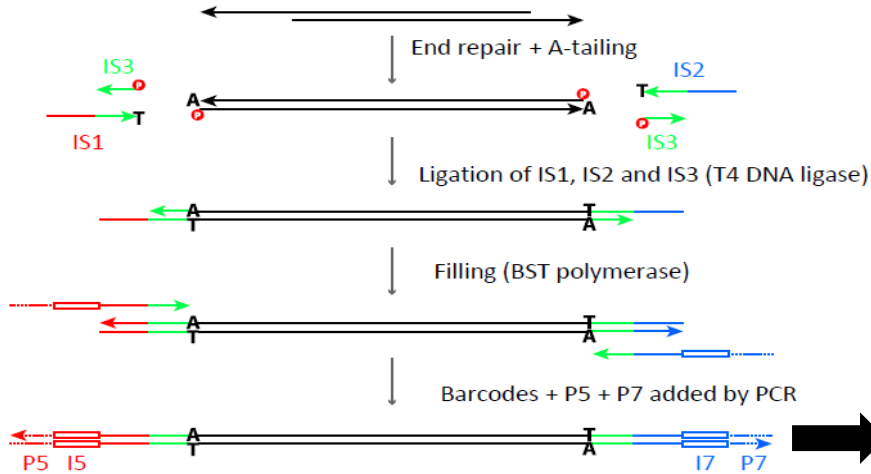
281 specimens:

239 ingroup

42 outgroup

Matterials and Methods

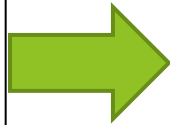
Library construction:



(Kircher 2011)



281 specimens:
239 ingroup
42 outgroup



281 Libraries

Matterials and Methods

Baits design:

Newly sequenced genomes



Bemangidia lowryi
81 million reads
20x – 40x



C. delphinense
51 million reads
2x – 20x



281 specimens:
239 ingroup
42 outgroup

281 Libraries

Matterials and Methods

Baits design:

GenBank transcriptome

Newly sequenced genomes



Manilkara zapota

Bemangidia lowryi

C. delphinense

81 million reads

20x – 40x

51 million reads

2x – 20x



281 specimens:

239 ingroup

42 outgroup



281 Libraries



Matterials and Methods

Baits design:

GenBank transcriptome

Newly sequenced genomes



Manilkara zapota

Bemangidia lowryi

C. delphinense

81 million reads

20x – 40x

51 million reads

2x – 20x

Baits design

Baits: small DNA sequences complementary to a locus that allow us to capture these locus from a genomic DNA solution by hybridization

Baits designed for:

227 microsatellites → Species complexes

532 genes from *Tseboneae* → Species level

262 genes from Angiosperms → Suprageneric level
(Johnson *et al.* 2018)



Baits for 1020 loci:

793 genes

227 microsatellites

281 Libraries

281 specimens:

239 ingroup

42 outgroup



Camille Christe

Matterials and Methods

Gene Capture

GenBank transcriptome



Manilkara zapota



Newly sequenced genomes

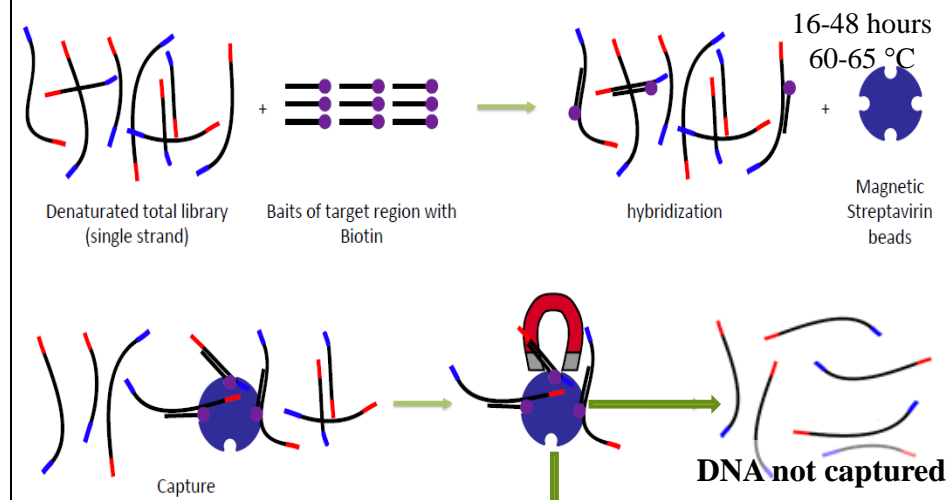


Bemangidia lowryi
81 million reads
20x – 40x



C. delphinense
51 million reads
2x – 20x

Gene Capture



Baits design



Baits for 1020 loci:

793 genes
227 microsatellites

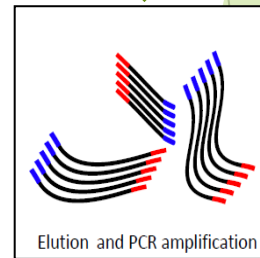
281 Libraries

Gene Capture

281 spécimens:

239 ingroup
42 outgroup

Pooled in 7 tubes according to size and baits specificity



Captured genes

Matterials and Methods

Sequencing

GenBank transcriptome



Manilkara zapota



Newly sequenced genomes

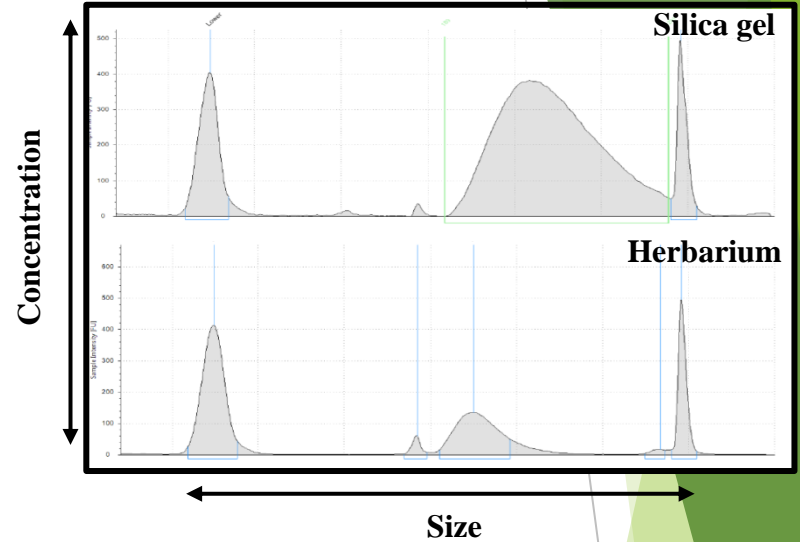


Bemangidia lowryi
81 million reads
20x – 40x



C. delphinense
51 million reads
2x – 20x

Example of tubes sent to sequencing



Baits design



Baits for 1020 loci:

793 genes
227 microsatellites

281 Libraries

Gene Capture

Captured loci

281 spécimens:

239 ingroup
42 outgroup

Pooled in 5 tubes
according to size

Pooled in 2 tubes
according to size

Illumina
sequencing

Unicopy genes

Multicopy genes

Matterials and Methods

Sequencing

GenBank transcriptome

Newly sequenced genomes



Manilkara zapota

Bemangidia lowryi

C. delphinense

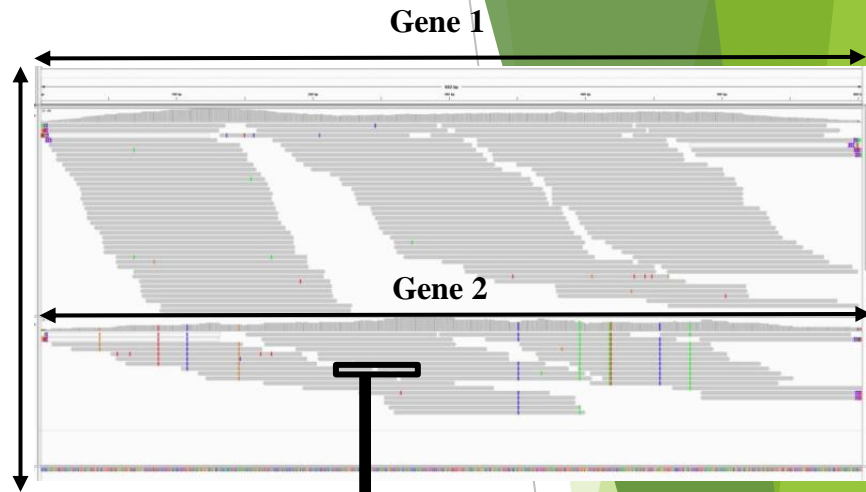
81 million reads

20x – 40x

51 million reads

2x – 20x

Number of copies



Baits design



Baits for 1020 loci:

793 genes

227 microsatellites

281 spécimens:

239 ingroup

42 outgroup

281 Libraries

Gene Capture

Captured loci

Pooled in 5 tubes
according to size

Pooled in 2 tubes
according to size

Illumina
sequencing

Unicopy genes

Multicopy genes

Sequences for
phylogenomic analyses

```

CTCACAATCCCTACACTCGGTAAATGAATGGAAACAGTAAGTTCCTCAGTCCCCATATATAAATATA
CTCACAATCCCTACACTCGGTAAATGAATGGAAACAGTAAGTTCCTCAATCCCCATATATAAATATA
GGTGCACAATGCCATGTGAAGCAATGGCTGGCTCTAAAGGACCCAGAGGCTGCTGGTCTTTCTTTTGT
GGTGCACAATGCCATGTGAAGCAATGGCTGGCTCTAAAGGACCCAGAGGCTGCTGGTCTTTCTTTTGT
TGAATGACAATATAAATAACCCATTTTCAAGATCAAGGTAACATCACTCAACATGCATCGCTCTTTTACAT
TGAATGACAATATAGATAACCCATTTTCAAGATCAAGGTAACATCACTCAACATGCATCGCTCTTTTACAT
    
```

Results

Gene Capture has been very efficient **➡** Storing method and collection year had not a strong impact



Fig. 2, Number of obtained genes according to the kind of sampling storage (silica-gel or herbarium) and collection year. Values in the upper part indicate the number of analysed samples per year, with the number of failed specimens between brackets.

Results

Gene Capture worked well even in other subfamilies

*= Specimen used for baits design

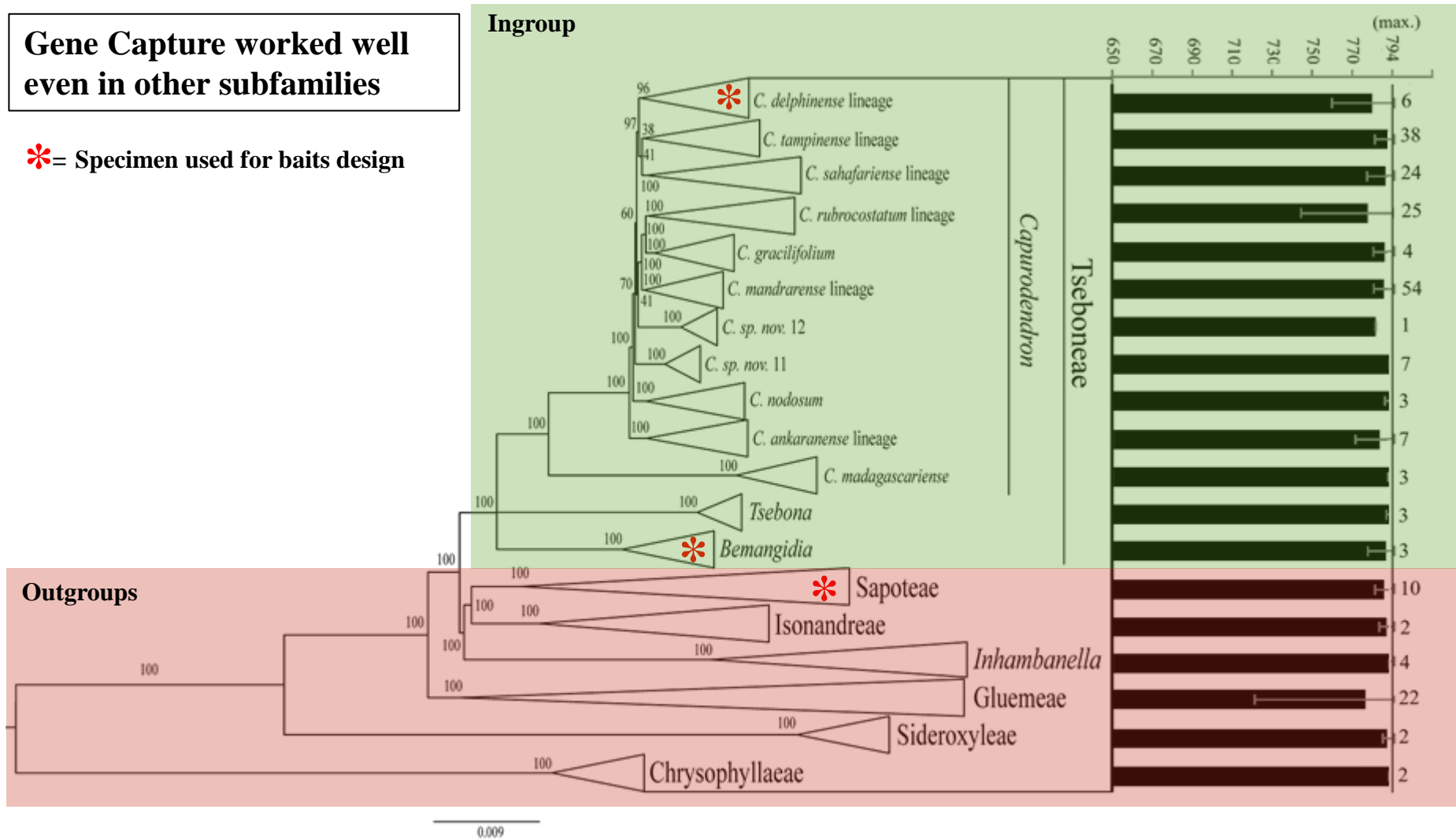
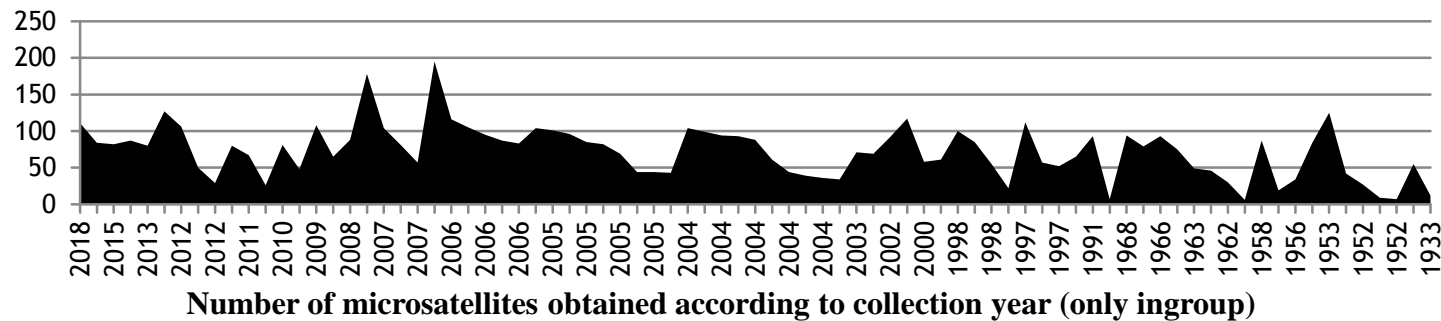
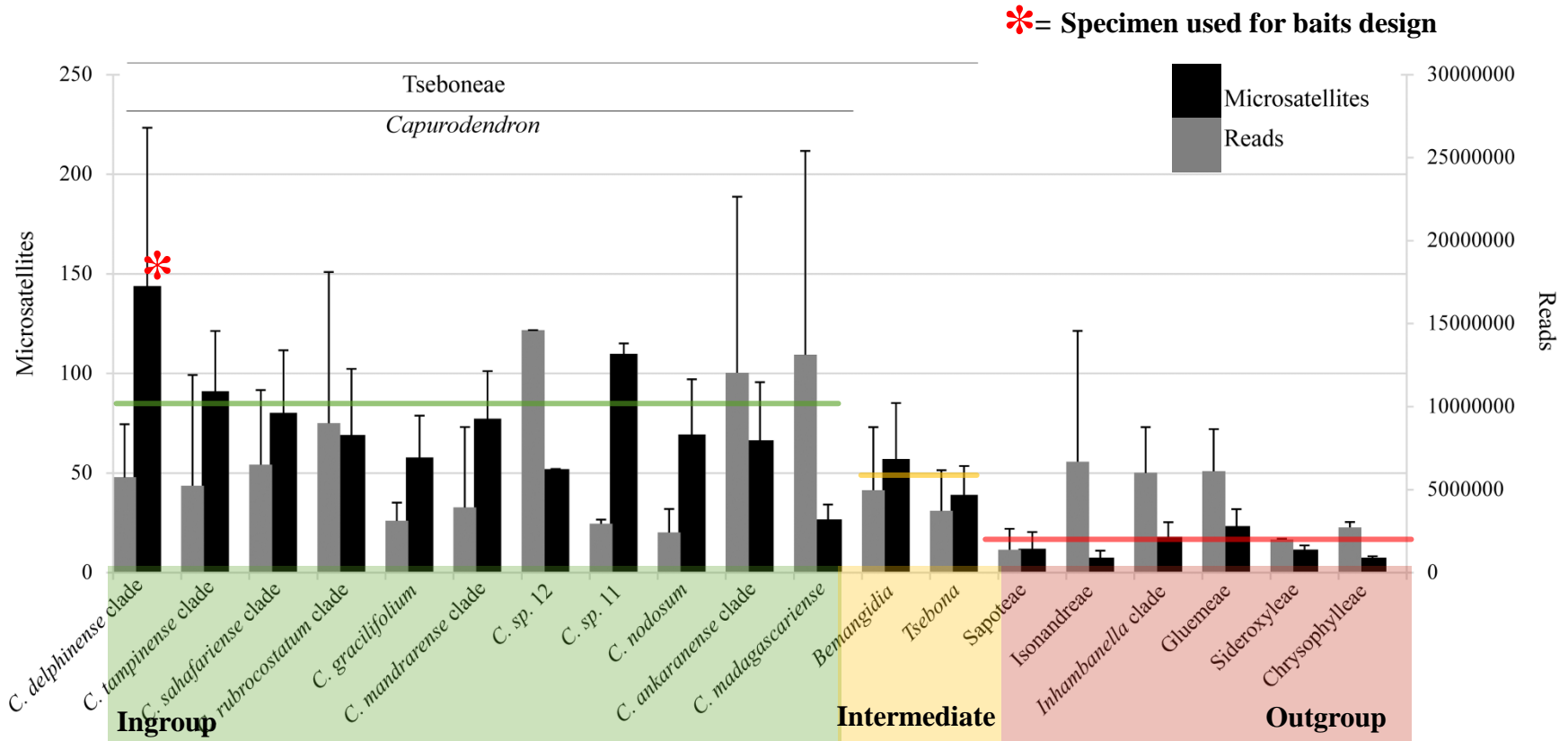


Fig.1 Maximum likelihood tree of 192192 SNPs from 519 coding genes and 444 OTUs (222 specimens). The major clades have been collapsed at tribe, genus or infrageneric lineage levels. Bars in the right margin indicate the average number of captured genes per clade and their standard deviation. The number of specimens per clade is indicated at the right margin.

Results

Microsatellite Capture efficiency is low \longrightarrow Independent of the number of reads / collection year



Results

Microsatellite Capture efficiency is low \longrightarrow Strongly dependent of phylogenetic proximity

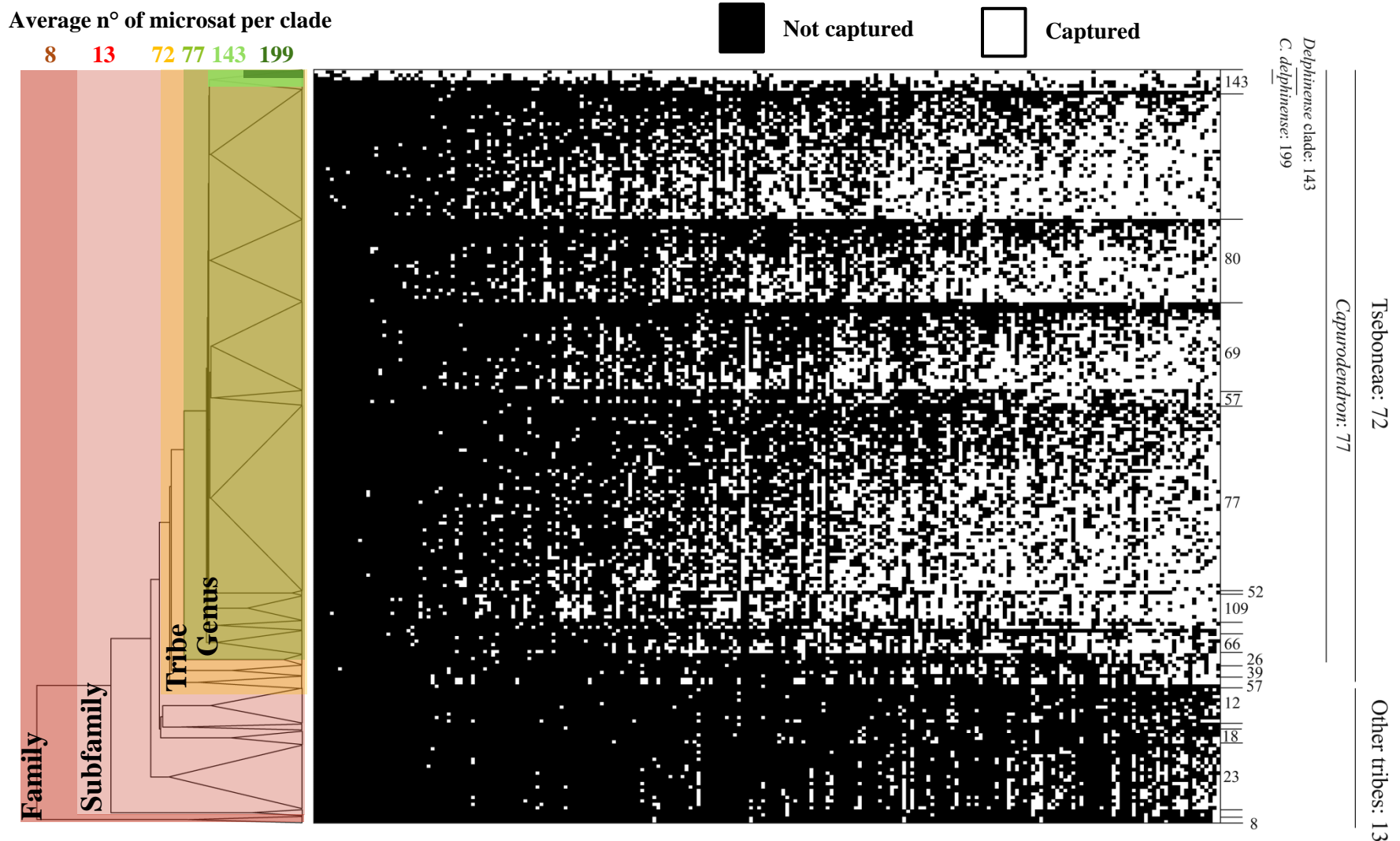
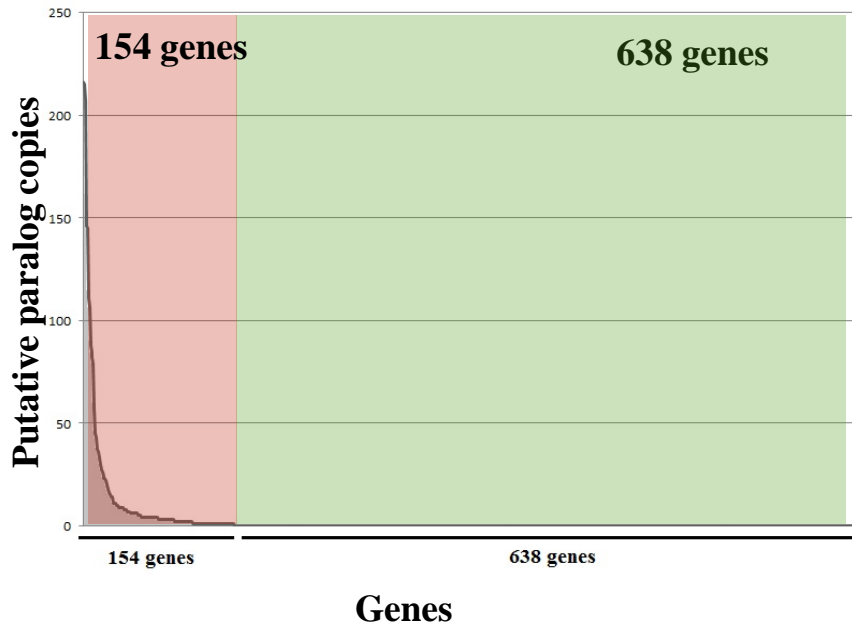


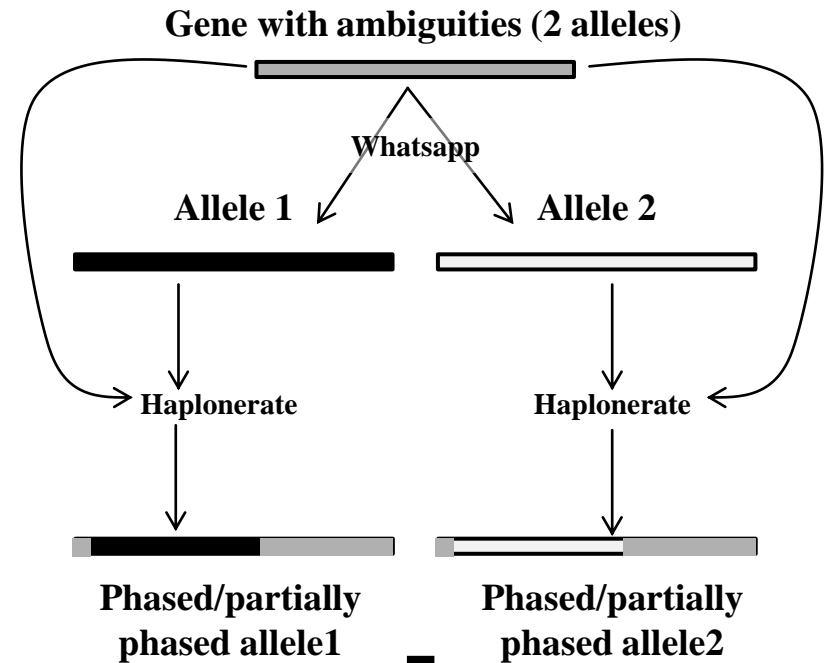
Fig. 4. Matrix showing failed microsatellites (black squares) and obtained (white squares) for each specimen and phylogenetic lineages. Numbers in the right margin indicate the average number of microsatellites successfully obtained.

Results

Gene selection



Phasing



Phylogeny

Cohalescence (BEAST):

-Sequences -34 genes
-444 OTUs -110574 bp
-72576 computing hours

Same tree topology

Maximum Likelihood (RaxML):

-SNPs -519 genes
-444 OTUs -192000bp
-4838 computing hours

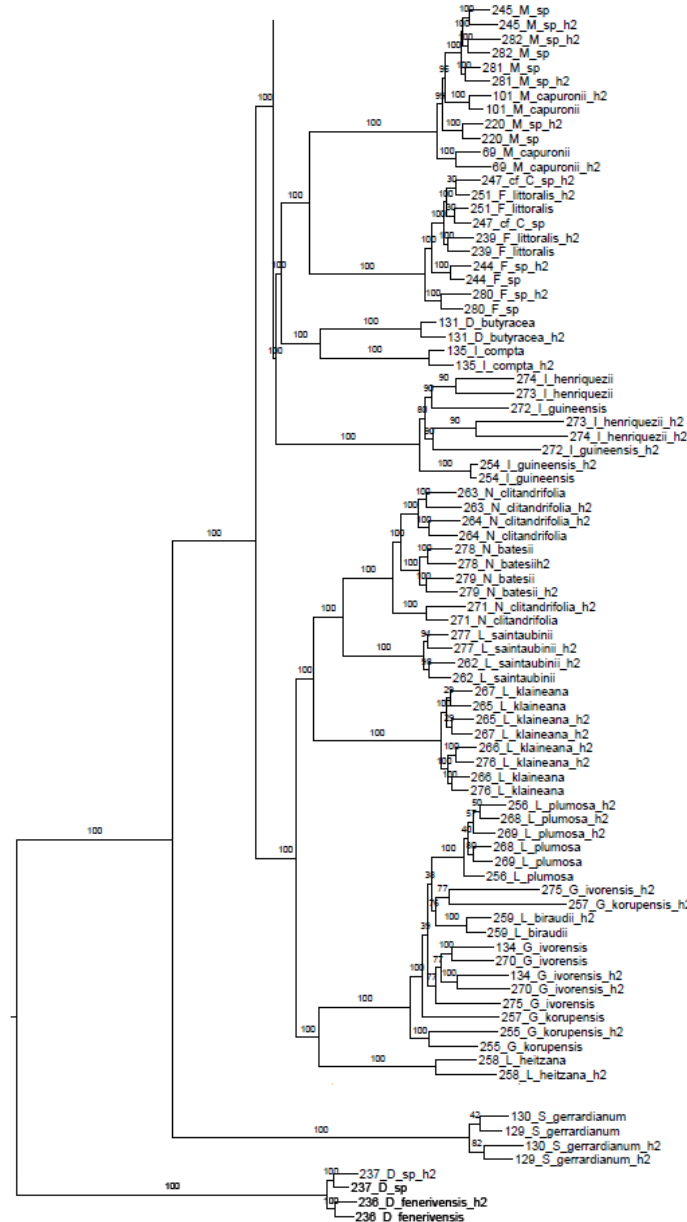
Results

Baits worked well in all the tribes



New projects has been started using the same baits

Tseboneae



Tseboneae

Sapoteae

Isonandreae

Inhambanella

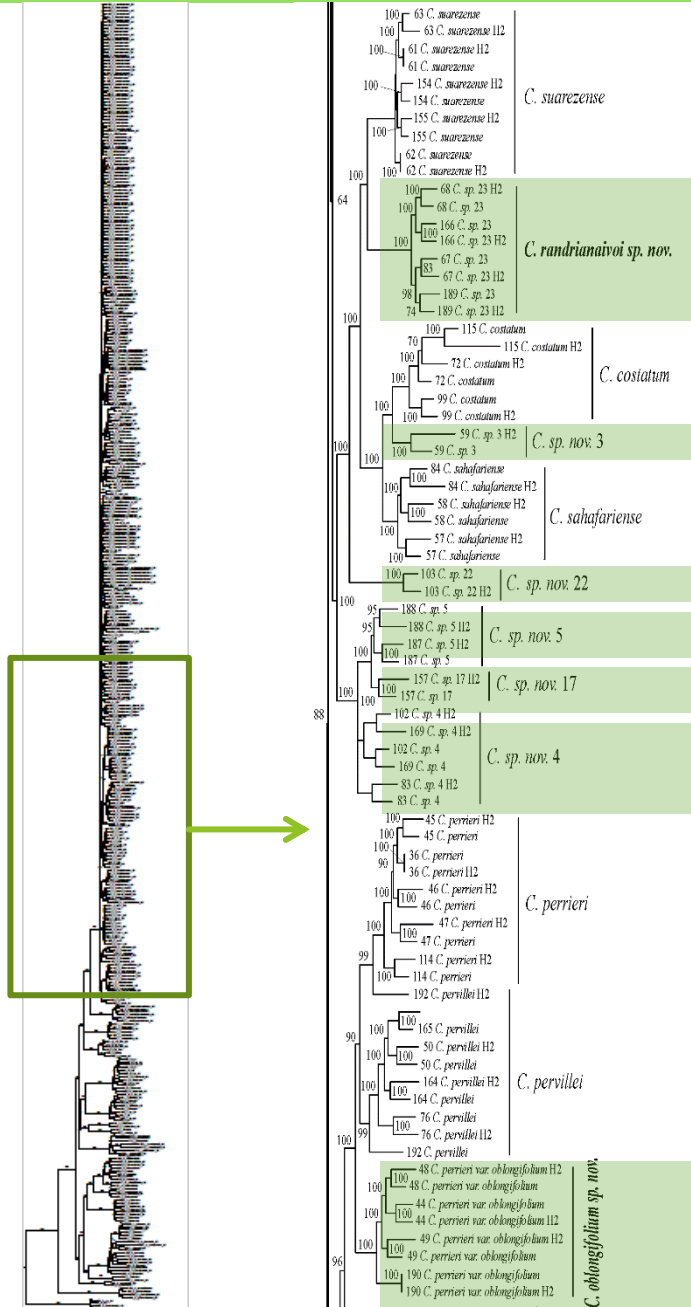
Gluemeae

Sideroxyleae

Chrysophylleae

RaxML tree 519 genes

Results



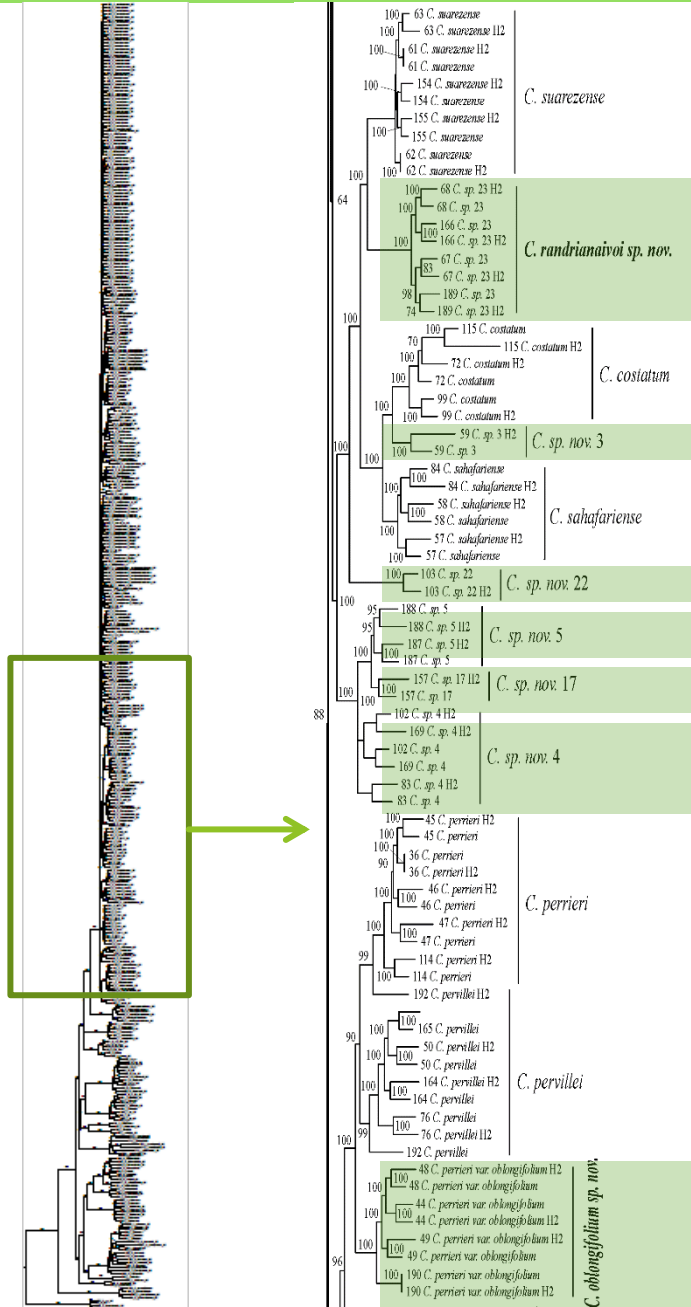
~ 49 clades/morphologies candidates for new species

Only 26 species described

23 new species?

= new species

Results



Described species

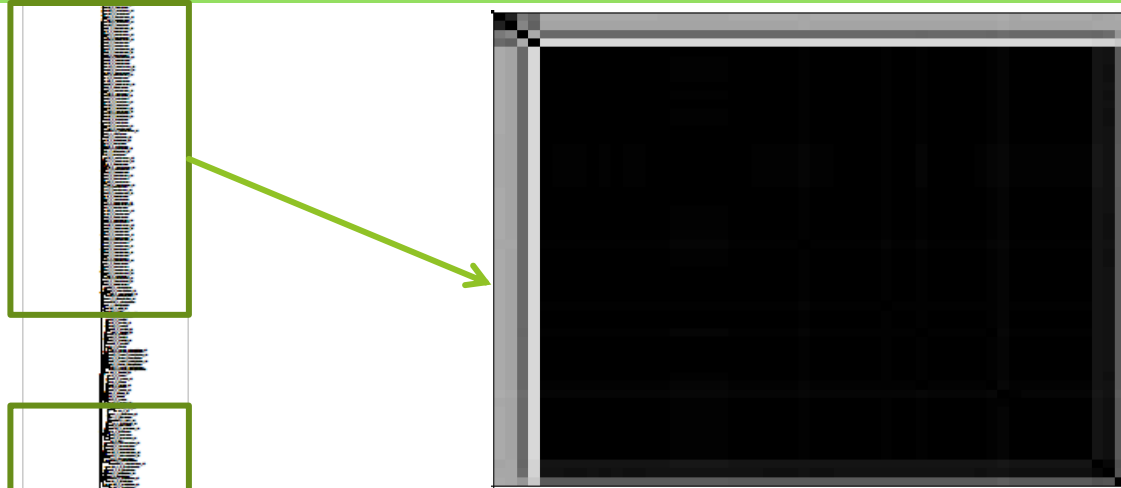


Undescribed species

= new species



Results



Arid species complex



Eastern species complex



Main Conclusions

- **The Gene Capture has been very efficient.**
- **Designed baits can be applied to all Sapotaceae family.**
- **Microsatellite capture only works well in the reference species.**
- ***Tseboneae* tribe may contain up to 23 undescribed species.**

Aknowledgements



Yamama Naciri



Richard Randrianaivo

Aina Randriarisoa

Carlos Galan Boluda



Camille Christe



Laurent Gautier

Thank you!



Contact: Carlos.g.boluda@gmail.com